

High-Performance Caged Nitric Oxide: A New Molecular Design, Synthesis, and Photochemical Reaction

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Endogenous nitric oxide is an important messenger molecule in both the brain¹ and blood vessels² and acts as a cytotoxic agent in the immune system.³ Since Kaplan reported the photolytic release of adenosine 5'-triphosphate (ATP) from a protected derivative that has no bioactivity, i.e., ATP *o*-nitrobenzyl ester,⁴ these photosensitive precursors, called caged compounds, have become an important tool in the biosciences, especially in exploring signal transductions in living systems.^{1b,5,6}

The great majority of known caged compounds are *o*-nitrobenzyl esters of desired bioactive molecules.⁵ Soon after we started this work, Makings and Tsien reported *o*-nitrobenzyl ester caged nitric oxides (NOs).⁷ The quantum yields of NO formation (Φ_{NO}) for their photochemical uncaging reaction were very poor (0.02–0.05) and NO chemical yields were only 30–54%.⁷ The efficiency of $\text{K}_2\text{Ru}(\text{NO})\text{Cl}_5$, used as a caged NO by Murphy et al., is also very low.⁶ Here, we report an extremely high-performance caged NO having a new molecular design.

The following three factors were considered in our molecular design of caged NO: (a) Because NO is a free radical, two NO molecules must be produced from a single caged NO molecule so that the remaining cage counterpart is a nonradical species after the photochemical reaction to produce the NO (uncaging). (b) Uncaging proceeds in a high Φ_{NO} . To achieve high Φ_{NO} , the weaker the bond between the NO and the cage counterpart in the caged NO, the better. If the bond is very weak, however, thermal uncaging occurs even at room temperature.⁸ (c) The caged NO requires a large molar extinction coefficient.

Fulfilling the above requirements, we designed *N,N'*-bis-(carboxymethyl)-*N,N'*-dinitroso-*p*-phenylenediamine (**I_A**) as a water-soluble caged NO and *N,N'*-dimethyl-*N,N'*-dinitroso-*p*-phenylenediamine (**I_B**) as a lipid-soluble caged NO.⁹ The homolysis of one N–N bond may induce homolysis of the second N–N bond, producing the second NO either stepwise or concerted (Scheme 1). The driving force of the expected reaction is the formation of the two stable free radicals and

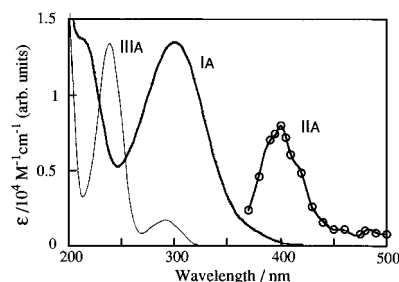
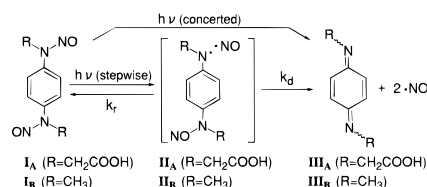
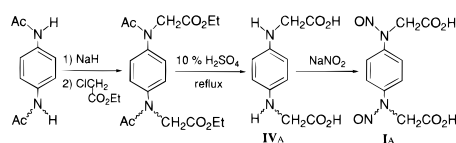


Figure 1. Electronic spectra. **I_A**: in 0.1 M sodium phosphate buffer, pH 7.4. **II_A**: Transient spectrum obtained by LFP of **I_A** in 0.1 M sodium phosphate buffer, pH 7.4. **III_A**: in 0.1% aqueous phosphoric acid.

Scheme 1



Scheme 2



formation of new C–N π -bondings in the broken-cage residue (**III**). Although homolytic cleavage of the N–N bond in mono-*N*-nitroso compounds occurs under irradiation, the radical pairs recombine, providing a starting material or a *C*-nitroso compound.^{10,11}

I_A was synthesized by the pathways shown in Scheme 2.^{12a} **I_B** was prepared by the nitrosation of *N,N'*-dimethyl-*p*-phenylenediamine (**IV_B**) with nitrous acid.^{12b} Caged NOs have strong absorption bands in the UV region (**I_A** λ_{max} = 300 nm (ϵ = 13.5 mM⁻¹ cm⁻¹) in a phosphate buffer of pH 7.4 (Figure 1); **I_B** λ_{max} = 299 nm (ϵ = 14.05 mM⁻¹ cm⁻¹) in methanol). Without light, a solution of **I_A** in a buffer of pH 7.4 at 37 °C initiated no reaction.

HPLC analysis of the reaction mixture after partial photolysis yielded only one peak besides the peak of the remaining starting **I_A** under different conditions. The broken cage residue of **I_A** was identified as **III_A** by comparison with the retention time and electronic spectrum (λ_{max} 290 nm) of the authentic sample prepared by the oxidation of amino acid **IV_A** with potassium ferricyanate (Scheme 3).¹³

Inter- and intracellular NO-mediated signal transductions are initiated by the coordination of NO with guanylate cyclase, a heme enzyme.¹⁴ To mimic this, **I_A** was photolyzed in a phosphate buffer solution of pH 7.4 containing mesotetrakis-(*p*-sulfonatophenyl)porphyrinocobalt (TPPSCo), a heme model,

(9) In the distribution equilibrium experiments between benzene and 0.1 M sodium phosphate buffer solution at pH 7.4, distribution ratios for **I_A** (K_{I_A}) and **I_B** (K_{I_B}) were found to be $K_{\text{I}_A} < 10^{-3}$ and $K_{\text{I}_B} > 5 \times 10^2$, where $K_{\text{I}} = [\text{I}]_{\text{benzene}}/[\text{I}]_{\text{water}}$.

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(12) (a) For **I_A**: mp 145–6 °C (dec). Anal. Calcd for C₁₀H₁₀N₂O₆: C, 42.55; H, 3.55; N, 19.86. Found: C, 42.49; H, 3.62; N, 19.55. (b) For **I_B**: mp 120–1 °C. Anal. Calcd for C₈H₁₀N₂O₂: C, 49.57; H, 5.18; N, 28.83. Found: C, 49.29; H, 5.20; N, 28.63.

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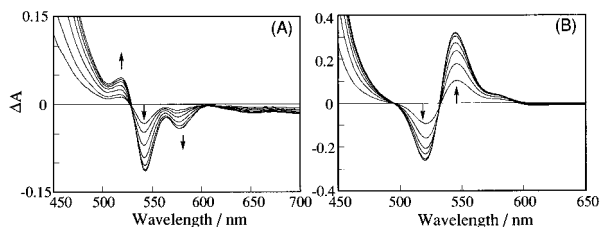


Figure 2. Difference spectra from photolysis of **I** in the presence of porphinatocobalt. (A) Photolysis of **I_A**: [**I_A**]₀ = 1.11 mM, [TPPSCo]₀ = 73.8 μM in 0.1 M phosphate buffer of pH 7.4; *t* = 0, 2, 4, 6, 8, 10, and 12 min. (B) Photolysis of **I_B**: [**I_B**]₀ = 0.78 mM, [TPPCo]₀ = 70 μM in THF; *t* = 0, 0.5, 1, 1.5, 3, 4, and 5 min.

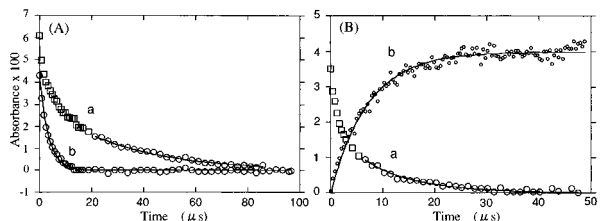
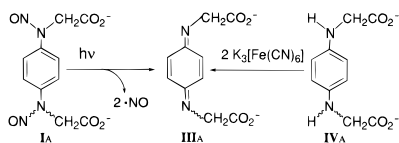


Figure 3. (A) Time courses for transient absorption at 405 nm in LFP of 69.6 μM **I_A** in pH 7.4 sodium phosphate buffer solutions. (a) Under Ar. (b) Under NO. (B) LFP of **I_B** in benzene under Ar. (a) Time course for transient absorption at 405 nm, [**I_B**]₀ = 63.6 μM. (b) Time course for raising absorption of TPPCo(NO) at 546 nm in the presence of 55 μM TPPCo, [**I_B**]₀ = 14.7 μM. All solid lines are first-order kinetics simulation curves.

Scheme 3



in an evacuated, sealed UV cuvette. **I_B** was uncaged in benzene containing mesotetraphenylporphinatocobalt (TPPCo). The spectrum change in photolysis (Figure 2) shows difference spectra obtained by subtracting the initial spectrum from those recorded at different photolytic uncaging time intervals. Difference spectra are identical to those obtained using authentic TPPSCo(NO) and TPPCo(NO) prepared by reacting TPPSCo and TPPCo with commercially obtained NO gas (λ_{\max} 542 nm, λ_{\min} 519 nm for TPPSCo(NO) and λ_{\max} 545 nm, λ_{\min} 520 nm¹⁵ for TPPCo(NO)). The photolytic reactions of caged NOs are clean (Figure 2).

When solutions of **I_A** in pH 7.4 sodium phosphate buffer and **I_B** in benzene were subjected to laser flash photolysis (LFP) with 308 nm wavelength and a 20 ns laser pulse at 23 °C, transient species having visible absorption band (Figure 1) appeared immediately. The time courses of the decreasing absorbance at 405 nm consist of fast phases (\square , Figure 3, trace a) and slow phases (\circ , Figure 3, trace a). Slow phases obeyed the first-order kinetic equation well ($-$, Figure 3) to yield $k_d = 2.96 \times 10^4 \text{ s}^{-1}$ for **I_A** in water and $k_d = 2.15 \times 10^5 \text{ s}^{-1}$ for **I_B** in benzene. Molecular oxygen did not alter the lifetime of the transient species, but NO accelerated decay. In a NO-saturated aqueous solution, the slow phase disappeared and the entire decay curve fitted a first-order kinetic equation with $k_{\text{obsd}} = 2.69 \times 10^5 \text{ s}^{-1}$ (Figure 3A, trace b). These observations and the NO trapping experiments, below, with TPPCo in benzene

(13) UV spectrum for **III_A** (Figure 1) was recorded by a multichannel detector for HPLC at the height of HPLC peaks. Quinonimines are unstable, so **III_A** was consecutively converted to secondary products. Since we were not interested in the reactions of **III_A**, we did not further examine secondary products.

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reveal that the photolysis of **I** occurs stepwise via radical intermediate **II**, which disappears via elimination of the second NO and recombination to regenerate **I**.

According to this mechanism, **II** disappears via the kinetic equation $-d[\mathbf{II}]/dt = (k_r[\text{NO}] + k_d)[\mathbf{II}]$. In the presence of a large excess NO, the bimolecular reaction predominates over the unimolecular NO extrusion reaction of **II** to yield $k_{\text{obsd}} = k_r[\text{NO}]$. From the solubility of NO at 23 °C (1.95 mM)¹⁶ and the k_{obsd} value, the second-order rate constant (k_r) for recombination of **II_A** and NO is calculated to be $1.38 \times 10^8 \text{ M}^{-1} \text{ s}^{-1}$. Since concentrations of caged NOs are 10^{-8} – 10^{-6} M in biological applications, the bimolecular recombination of NO and **II** should be negligible in practical use.

When the LFP of **I_B** benzene solutions having a constant initial concentration in the presence of a large excess TPPCo was performed, time courses of rising absorption at 546 nm due to the formation of TPPCo(NO) obeyed the first-order kinetic equation (Figure 3B, trace b). Pseudo-first-order rate constants correlated linearly with [TPPCo] to obtain the second-order rate constant for the binding of NO and TPPCo ($2.80 \times 10^9 \text{ M}^{-1} \text{ s}^{-1}$), which agrees with previously reported $7 \times 10^8 \text{ M}^{-1} \text{ s}^{-1}$ in 2-methyltetrahydrofuran.¹⁵ When the benzophenone T–T band absorbance ($\epsilon = 10\,300$) at 532.5 nm was used as a standard,¹⁷ the quantum yield for the formation of TPPCo(NO), $\Phi_{\text{TPPCo(NO)}}$, was determined to be 1.97 ± 0.33 from LFP with [**I_B**] = 7.2 μM and [TPPCo] = 60 μM. In low TPPCo concentration, not all NOs could be captured by TPPCo. As was predicted in our molecular design, a single photon generates almost two NO molecules from these caged NOs.

To further substantiate uncaging efficiency, a solution of **I_A** (21.3 μM) in a 0.1 M sodium phosphate buffer solution at pH 7.4 was irradiated until all **I_A** was photolyzed under Ar in the presence of 0.23 mM 2-(4-carboxyphenyl)-3,3,4,4-tetramethylimidazoline-1-oxyl 3-oxide (carboxy-PTIO), which reacts with the NO fading blue (λ_{\max} 561 nm, $\epsilon = 970 \text{ M}^{-1} \text{ cm}^{-1}$).¹⁸ Light having a 300 ± 0.5 nm wavelength was provided from a 500 W xenon lamp through a monochromator. One mole of **I_A** was found to consume 1.98 ± 0.01 mol of carboxy-PTIO. Similarly, in irradiation of a tetrahydrofuran solution containing **I_B** and large excess amounts of 2-phenyl-3,3,4,4-tetramethylimidazole-1-oxyl 3-oxide (PTIO), 1 mol of **I_B** reacted with 2.05 ± 0.10 mol of PTIO. These observations reveal that 1 mol of caged NOs quantitatively yields 2 mol of NO upon uncaging.

The values of the quantum yield consuming PTIO, Φ_{PTIO} , were determined using a potassium ferrioxalate actinometer.¹⁹ Values of the experimentally observed Φ_{PTIO} in uncaging were 1.87 ± 0.08 with 300 nm light, 1.6 ± 0.1 with 350 nm light for **I_A**, and 2.02 ± 0.03 for **I_B** with 300 nm light.²⁰ The quality of the caged compound is expressed by $\epsilon\Phi$. The $\epsilon\Phi$ values of **I_A** were $2.7 \times 10^4 \text{ M}^{-1} \text{ cm}^{-1}$ with 300 nm light and $6 \times 10^3 \text{ M}^{-1} \text{ cm}^{-1}$ with 350 nm light. This is much greater than those for reported *o*-nitrobenzyl ester caged NOs ($\epsilon\Phi = 15$ – $70 \text{ M}^{-1} \text{ cm}^{-1}$).⁷

The assay of caged NOs on rat aorta vasorelaxation is currently underway at this laboratory. Preliminary results reveal that caged NOs are very high-performance and have no measurable toxicity.

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(20) During the 25 ns interval after laser pulse irradiation of an aqueous **I_A** solution, a sharp decay in absorbance accounting for several percent of all **II_A** decay occurred. This may be attributed to the recombination of radicals pair in the solvent cage to yield **I_A**. No such phenomenon was observed, however, in LFP of **I_B** in benzene. This may explain the somewhat lower quantum yield in water than benzene.